# ANTITUMOR AGENTS, 74.<sup>1</sup> BRUCEANOL-A AND -B, TWO NEW ANTILEUKEMIC QUASSINOIDS FROM BRUCEA ANTIDYSENTERICA

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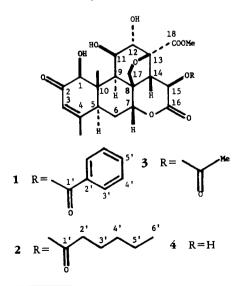
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Eight quassinoids, including the antileukemic compound bruceantin, which was in phase II clinical trial by the National Cancer Institute, have been isolated from the Ethiopian tree Brucea antidysenterica Mill. (Simaroubaceae) by Kupchan and associates (1). In a previous paper, we reported the isolation and structural elucidation of two new antileukemic quassinoid glycosides, bruceantinoside-A and -B, from B. antidysenterica (2). We now describe the isolation and characterization of two additional new antileukemic isobruceine type quassinoids, bruceanol-A (1) and -B (2), from this same plant.

## **RESULTS AND DISCUSSION**

Bruceanol-A (1) had the molecular formula  $C_{28}H_{30}O_{11}$  from both field de-



<sup>1</sup>For part 73, see H. Nozaki, H. Suzuki, T. Hirayama, R. Kasai, R.Y. Wu, and K.H. Lee, *Phytochemistry*, (in press).

sorption and hrms. Compound 1 showed the presence of hydroxyl (3540, 3450, and 1030 cm<sup>-1</sup>), phenyl (1600) and 1580 cm<sup>-1</sup>),  $\delta$ -lactone (1750 cm<sup>-1</sup>), ester (1730 cm<sup>-1</sup>), and  $\alpha$ ,  $\beta$ -unsaturated carbonyl (1675  $\text{cm}^{-1}$ ) groups in its ir spectrum. The <sup>1</sup>H-nmr spectrum of 1 is identical with that of isobruceine-B (3), except for the signals in the acid portion of the ester as well as the COOMe-13 and the H-15. These differences were observed in 1 as a phenyl ring [8 8.0-8.05 (2H), H-3' and 7.42-7.65 (3H), H-4' and H-5'] and in 3 as a methyl group [ $\delta$  2.08 (3H, s)]. In **1**, the signal for COOMe-13 ( $\delta$  3.51) was shifted upfield by 0.32 ppm, and that for H-15 ( $\delta$  6.49) was shifted downfield by 0.19 ppm compared to the corresponding signals of **3** ( $\delta$  3.83 for COOMe-13 and  $\delta$  6.30 for H-15) due to the anisotropic effect of the aromatic ring in 1 (Table 1).

Added confirmation for these assignments was obtained by a comparison of the <sup>13</sup>C-nmr spectra of **1** and **3** (Table 2). The carbon signals for both compounds were superimposable except for the differences in the above mentioned ester side chains. Thus, **1** contained the aromatic carbons at  $\delta$  128.7 (s, C-2'), 128.5 (d, C-3'), 130.0 (d, C-4'), and 133.8 (d, C-5'), whereas **3** possessed a quartet methyl group at  $\delta$  20.5 (C-2').

Bruceanol-B (2) was analyzed for  $C_{27}H_{36}O_{11}$ . Its ir spectrum was consistent with the presence of hydroxyl (3540, 3450, and 1030 cm<sup>-1</sup>),  $\delta$ -lactone (1750 cm<sup>-1</sup>), ester (1730 cm<sup>-1</sup>), and  $\alpha$ , $\beta$ -unsaturated carbonyl (1677 cm<sup>-1</sup>) groups. Compound 2 showed

also comparable <sup>1</sup>H-nmr (Table 1) and <sup>13</sup>C-nmr (Table 2) spectra to those of **3** except for the signals attributable to a hexanoyl group [ $\delta$  1.3 (m, H-3', H-4', and H-5') and 0.91 (t, J=7.0 Hz, H-6'), and  $\delta$  33.6 (t, C-2'), 31.2 (t, C-3'), 24.2 (t, C-4'), 22.2 (t, C-5'), and 13.8 (q, C-6')] in **2** and an acetyl [ $\delta$  2.08 (s) and  $\delta$  20.5 (q, C-2')] moiety in **3**.

Deacetylisobruceine-B (4) was obtained by alkaline hydrolysis of 1,2, and 3. Thus, 1 and 2 were structurally identical with 3 except for the slight differences in the C-15 ester side chain, in which 1 and 2 have a benzoate and an hexanoate, respectively, whereas 3 bears an acetate moiety. an internal standard. Mass spectra were recorded on a JEOL JMS-D100 (EI) and/or a Hitachi M-80 (EI and FD) instruments. Silica gel (Merck, type 60, 70-230 mesh was used for column chromatography, and precoated silica gel plates (Merck, 60 F-254, 0.25 and 2mm) were used for analytical and preparative tlc, respectively. Detection of components was made by use of a uv lamp. Hplc was performed on a Waters Associates Model ALC/GPC 244 liquid chromatograph using a  $\mu$ Bondapak C18 column (3.9 mm×300 mm) for analytical, and M and S PAK C18-A (20 mm×250 mm) for preparative purposes.

CHROMATOGRAPHY OF THE CHCl<sub>3</sub> FRAC-TIONS.—The crude CHCl<sub>3</sub> fraction (372 g), which is part of the CHCl<sub>3</sub> extract of the ground wood of *B. antidysenterica* (4228 lbs) reported previously (2), was subjected to column chromatography on silica gel (2 kg, 10 cm  $\times$  60 cm and

_	Bruceanol- $A(1)^a$	Bruceanol-B (2) <sup>b</sup>	Isobruceine-B (3) <sup>c</sup>	Deacetylisobruceine-B (4) <sup>d</sup>
H-1	4.32 s	4.28 br.s	4.26 s	4.85 br.s
H-3	6.13 br.s	6.1 br.s	6.11 br.s	6.13 br.s
H-7	4.8 m	4.75 br.s	4.75 m	5.54 t (4.9) <sup>e</sup>
<b>H-</b> 11	4.57 s	4.54 br.s	4.75 m	5.08 br.s
H-12	4.20 s	4.18 br.s	4.12 br.s	4.35 br.s
H-15	$6.49 d (13)^{e}$	6.29 d (13) <sup>e</sup>	6.30 d (13) <sup>e</sup>	6.12 d (12.2) <sup>e</sup>
OMe	3.51s	3.83 s	3.83 s	3.79 s
Me-4	1.98 s	1.96 s	1.95 s	1.72 s
<b>Me-10</b>	1.21s	1.19 s	1.20 s	1.44 s
2'		_	2.08 s	
3' 4'	8.0-8.05 (2H)	1.3 m		
5'	7.42-7.65(3H)			_
6'	-	0.91 t (7) <sup>e</sup>	-	

TABLE 1. <sup>1</sup>H-nmr Spectra of Quassinoids 1, 2, 3, and 4

<sup>a</sup>Measured at 200 MHz in CDCl<sub>3</sub>.

<sup>b</sup>Measured at 400 MHz in CDCl<sub>3</sub>.

<sup>c</sup>Measured at 400 MHz in CDCl<sub>3</sub> by Polonsky et al. (3).

<sup>d</sup>Measured at 400 MHz in C<sub>5</sub>D<sub>5</sub>N.

\*Coupling constant in Hz.

## **EXPERIMENTAL**

GENERAL EXPERIMENTAL PROCEDURES.— Melting points were determined on an MRK airbath-type melting point apparatus and were uncorrected. Specific rotations were obtained on a Yanagimoto OR-50 polarimeter (L=0.5 dm). Ir and uv spectra were recorded on a Hitachi 215 grating ir spectrometer and a Hitachi 320-S uv spectrometer, respectively. <sup>1</sup>H-nmr and <sup>13</sup>C-nmr spectra were determined on a JEOL JNM-FX 200 (200 MHz for <sup>1</sup>H nmr and 50.1 MHz for <sup>13</sup>C nmr) and/or a JEOL JNM-GX400 (400 MHz for <sup>1</sup>H nmr and 100 MHz for <sup>13</sup>C nmr) using TMS as eluted first with CHCl<sub>3</sub> and then with increasing amounts of MeOH in CHCl<sub>3</sub> to yield 28 fractions. Fraction 20 (31 g) was further subjected to column chromatography on silica gel (500 g, 4 cm×90 cm and eluted with EtOAc-Et<sub>2</sub>O, 1:1, v/v) to afford a pale yellow gum (6.7 g) which showed the presence of isobruceine-B (3), bruceanol-A (1), and bruceanol-B (2) on analytical hplc (MeOH-H<sub>2</sub>O, 1:1, v/v).

ISOLATION OF BRUCEANOL-A (1).—The pale yellow gum (6.7 g) was subjected to preparative hplc using a mixed solvent of MeOH-H<sub>2</sub>O (1:1, v/v) to give a colorless amorphous compound (1.1

	Bruceanol-A(1) <sup>a</sup>	Bruceanol-B(2) <sup>b,c</sup>	Isobruceine-B (3) <sup>d</sup>	Deacerylisobruceine-B (4) <sup>e</sup>
C-1	80.7 d	80.6 d	81.3 d	82.9 d
C-2	196.7 s	196.9 s	197.6s	198.4 s
C-3	124.3 d	124.3 d	124.5 d	125.0 d
C-4	163.0 s	162.9 s	162.6 s	163.2 s
C-5	43.5 d	43.5 d	43.4 d	43.9 d
C-6	28.6 t	28.5 t	28.2 d	28.6 t
<b>C-</b> 7	81.3 d	81.1 d	81.7 d	83.1 d
C-8	45.8 s	45.7 s	45.8 s	46.2 s
C-9	42.8 s	42.7 d	42.4 d	43.4 d
C-10	47.6s	47.5 s	47.7 s	48.4 s
C-11	72.7 d	72.5 d	74.3 d	75.2 d
C-12	75.8d	75.8 d	75.1d	76.9 d
C-13	82.9 s	82.8 s	81.7 s	82.6 s
C-14	51.4d	51.5 d	52.3 d	55.5 d
C-15	67.4d	66.5 d	67.8 d	66.3 d
C-16	166.7 s	166.9 s	167.6 s	172.8 s
<b>C-17</b>	73.3 t	73.3 t	73.0 t	74.0 t
C-18	172.0 s	172.4 s	169.5 s	173. <b>5 s</b>
OMe	52.9 q	52.8 q	49.8 q	52.3 g
Me-4	22.5 q	22.5 q	22.4 q	22.2 g
Me-10	11.6 g	11.5 q	11.3 g	11.4 g
C-1'	164.8 s	171.9 s	170.7 s	—
C-2′	128.7 s	33.6 t	20.5 q	—
C-3′	128.5 d	31.2 t		
C-4′	130.0 d	24.2 t	—	
C-5′	133.8 d	22.2 t		<u> </u>
C-6'		13.8 q	—	

TABLE 2. <sup>13</sup>C-nmr Spectra of Quassinoids 1, 2, 3, and 4

<sup>a</sup>Measured at 50.1 MHz in CDCl<sub>3</sub>.

<sup>b</sup>Measured at 100.40 MHz in CDCl<sub>3</sub>.

<sup>c</sup>The characterizations of sec-C was based on the INEPT (Insensitive Nuclei Enhanced by Polarization Transfer).

<sup>d</sup>Measured at 22.63 MHz in CDCl<sub>3</sub> by Polonsky et al. (3).

<sup>e</sup>Measured at 100.40 MHz in  $C_5D_5N$ .

g), which was eluted after isobruceine-B. This amorphous compound was further subjected to preparative hplc to yield pure bruceanol-A (1, colorless and amorphous, 0.000058% yield); mp 174-177° (from CH<sub>2</sub>Cl<sub>2</sub>/Et<sub>2</sub>O);  $\{\alpha\}^{25}D + 60^{\circ}$  (c 0.5, EtOH); uv  $\lambda$  max (EtOH) 232 nm ( $\epsilon$ 22400); The ir spectral data were discussed in the text. The <sup>1</sup>H-nmr and <sup>13</sup>C-nmr data were summarized in Table 1 and 2, respectively. Fdms m/z543 (M<sup>+</sup>+1); eims m/z 542.1853 (calcd. for C<sub>28</sub>H<sub>30</sub>O<sub>11</sub>: 542.1788, M<sup>+</sup>), 524,1657 (calcd. for C<sub>28</sub>H<sub>28</sub>O<sub>10</sub>: 524.1682, M<sup>+</sup>-H<sub>2</sub>O), 492.1418 (calcd. for C<sub>27</sub>H<sub>24</sub>O<sub>9</sub>: 492.1420, M<sup>+</sup>-MeOH), and 420.1438 (calcd. for C<sub>21</sub>H<sub>24</sub>O<sub>9</sub>: 420.1420, M<sup>+</sup>-C<sub>6</sub>H<sub>5</sub>COOH).

ISOLATION OF BRUCEANOL-B (2).—The pale yellow gum (6.7 g) was further eluted after the elution of bruceanol-A to afford a compound (0.54 g). This compound was purified by preparative hplc to yield pure bruceanol-B (2, colorless and amorphous, 0.000028% yield); mp 170-172° (from EtOAc/Et<sub>2</sub>O);  $[\alpha]^{25}D + 32°$  (c 0.5, MeOH); uv  $\lambda$  max (EtOH) 236 nm ( $\in$  8300); fdms m/z 537 (M<sup>+</sup>+1); Found: C, 60.49%; H, 6.74%. Calcd. for C<sub>27</sub>H<sub>36</sub>O<sub>11</sub>: C, 60.43%, H, 6.76%.

ISOLATION OF ISOBRUCEINE-B (3).—Isobruceine-B (3) was obtained as a colorless and amorphous compound (0.1 g) by the preparative hplc of the foregoing pale yellow gum (6.7 g). It was also obtained in 4% yield from fraction 21 (88 g) in the same manner as described above. This compound was identified by comparing its <sup>1</sup>H-nmr (400 MHz) spectrum (Table 1) with that presented by Polonsky (3).

HYDROLYSIS OF ISOBRUCEINE-B (**3**).—To a solution of isobruceine-B (**3**, 25 mg) in MeOH (1.5 ml) was added 1N NaOH (0.45 ml). After stirring at  $-15^{\circ}$  for 17 min, the reaction mixture was neutralized with 1N HCl (0.8 ml) and evaporated in vacuo to give a residue. The residue was subjected to preparative tlc to afford pure deacetylisobruceine-B (**4**) as colorless needles (8.2

mg); mp 279-282° (from ErOAc/Et<sub>2</sub>O); ir (KBr) 3400, 1735 and 1670 cm<sup>-1</sup>; eims m/z 438.1538 (M<sup>+</sup>; calcd. for C<sub>21</sub>H<sub>26</sub>O<sub>10</sub>: 438.1526); Found: C, 57.49%; H, 5.99%. Calcd. for C<sub>21</sub>H<sub>26</sub>O<sub>10</sub>: C, 57.53%; H, 5.98%.

HYDROLYSIS OF BRUCEANOL-A (1).— Bruceanol-A (1, 46.6 mg) was hydrolyzed and worked up in the same manner described above to give colorless needles (6.2 mg) identified as deacetylisobruceine-B ( $\stackrel{\bullet}{4}$ ) by comparing its <sup>1</sup>Hnmr (400 MHz, C<sub>5</sub>D<sub>5</sub>N) spectrum with that of an authentic sample derived from the hydrolyzed product of isobruceine-B.

HYDROLYSIS OF BRUCEANOL-B (2).— Bruceanol-B (2, 25 mg) was hydrolyzed and worked up in the same manner as described above to yield colorless needles (7.2 mg) identified as deacetylisobruceine-B (4) by a direct <sup>1</sup>H-nmr comparison with an authentic sample.

BIOLOGICAL ACTIVITY.—In vivo antitumor assay was carried out according to a standard National Cancer Institute procedure described in the literature (4). Bruceanol-A (1) and -B (2) showed significant (T/C $\geq$ 120%) antileukemic activity in vivo against P-388 lymphocytic leukemia (3-day dosing) at T/C=130% (0.5 mg/kg), 129% (1 mg/kg) and 134% (2 mg/kg), and 123% (0.5 mg/kg), respectively. The control, 5-fluorouracil, had T/C=135% (200 mg/kg, 1-day dosing).

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#### LITERATURE CITED

- S.M. Kupchan, R.W. Britton, J.A. Lacadie, M.F. Ziegler, and C.W. Sigel, J. Org. Chem., 40, 648 (1975).
- M. Okano, K.H. Lee, I.H. Hall, and F.E. Boettner, J. Nat. Prod., 44, 470 (1981).
- C. Moretti, J. Polonsky, M. Vuilhorgne, and T. Prange, *Tetrabedron Lett.*, 23, 647 (1982).
- R.I. Geran, N.H. Greenberg, M.M. Mac-Donald, A.M. Schumacher, and B.J. Abbott, *Cancer Chemother. Rep., Part 3*, 3, 1 (1972).

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